

# Package ‘SimPhe’

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**Type** Package

**Title** Tools to Simulate Phenotype(s) with Epistatic Interaction

**Version** 0.2.0

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**Description** Provides functions to simulate single or multiple, independent or correlated phenotype(s) with additive, dominance effects and their interactions. Also includes functions to generate phenotype(s) with specific heritability. Flexible and user-friendly options for simulation.

**BugReports** <https://github.com/beibeiJ/SimPhe/issues/new>

**URL** <https://github.com/beibeiJ/SimPhe>

**License** GPL (>= 2)

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---

allele.freq	<i>Frequencies of SNPs.</i>
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---

## Description

A dataset containing sample allele frequencies of SNPs. allele.freq.

## Usage

```
data(allele.freq)
```

## Format

A data frame with 6 rows and 3 columns(variables):

**SNP** name of SNP

**major.frequency** frequency of major allele

**minor.frequency** frequency of minor allele

---

build.cor.phe                    *Build correlated phenotypes*

---

**Description**

Build correlated phenotypes

**Usage**

```
build.cor.phe(pheno, corMtr, sdMtr = NULL, margin = 2, ...)
```

**Arguments**

pheno	a matrix or dataframe with the phenotypic information.
corMtr	a correlation matrix.
sdMtr	a matrix with the standard deviation, e.g., if the number of dimensions is 2, then it is $\begin{pmatrix} \sigma_1 & 0 \\ 0 & \sigma_2 \end{pmatrix}$ . If it is NULL (default), generate it based on the data from pheno.
margin	a vector giving the subscript which the function will be applied over. E.g., for a matrix 1 indicates rows, 2 (default) indicates columns. Where pheno has named dimnames, it can be a character vector selecting dimension names.
...	not used.

**Value**

a matrix with correlated phenotypes.

**Author(s)**

Beibei Jiang <beibei\_jiang@psych.mpg.de>

**Examples**

```
x1 <- rnorm(4000, mean = 5, sd = 10)
x2 <- rnorm(4000, mean = 10, sd = 30)
x <- matrix(cbind(x1, x2), ncol = 2)

# test original correlation
cor.test(x[, 1], x[, 2])

# correlation matrix
corM <- matrix(c(1, 0.6, 0.6, 1), ncol = 2)

# standard deviation matrix
sdM <- matrix(c(10, 0, 0, 30), ncol = 2)

# build correlation
```

```
x.new <- build.cor.phe(x, corM, sdM)

# check mean and standard deviation of new data set
apply(x.new, 2, mean)
apply(x.new, 2, sd)

# test correlation
cor.test(x.new[, 1], x.new[, 2])
```

---

build.sd.matrix      *build a matrix with standard deviation*

---

## Description

Build a matrix with standard deviation.

## Usage

```
build.sd.matrix(x, margin = 2, ...)
```

## Arguments

x	a matrix or dataframe.
margin	an integer specifying the dimension that standard deviation will be computed for. 1 indicates rows, 2 indicates columns. Default is 2.
...	not used

## Value

a matrix with the standard deviation. If the number of dimensions is 2, then it is  $\begin{pmatrix} \sigma_1 & 0 \\ 0 & \sigma_2 \end{pmatrix}$ .

## Author(s)

Beibei Jiang <beibei\_jiang@psych.mpg.de> and Benno Pütz <puetz@psych.mpg.de>

## Examples

```
x1 <- rnorm(4000, mean = 5, sd = 10)
x2 <- rnorm(4000, mean = 10, sd = 30)
x <- matrix(cbind(x1, x2), ncol = 2)
build.sd.matrix(x)
```

---

calc.gene.var	<i>Genetic Variance</i>
---------------	-------------------------

---

**Description**

Calculate the total genetic variance

**Usage**

```
calc.gene.var(gene.coef, freq, Dskim = 0, ...)
```

**Arguments**

gene.coef	a list with the coefficients of genetic effects. Each element includes 8 genetic parameters (regression coefficient) $\beta_{G_{w_t}}$ , $t$ in $(1, 2, \dots, 8)$
freq	a dataframe with the allele frequencies.
Dskim	the coefficient of linkage disequilibrium. Default is 0 (no LD).
...	not used.

**Details**

The genetic variance is calculated based on the genetic parameters  $\beta_{G_{w_t}}$ ,  $t$  in  $(1, 2, \dots, 8)$ , as described in the publications by Kao and Cockerham:

Kao CH, Zeng ZB. Modeling epistasis of quantitative trait loci using Cockerham's model. *Genetics*. 2002 Mar 1;160(3):1243-61.  
<http://www.genetics.org/content/160/3/1243.short>

Cockerham CC, Weir BS. Quadratic analyses of reciprocal crosses. *Biometrics*. 1977 Mar 1:187-203.  
<http://www.jstor.org/stable/2529312>

**Value**

genetic variance.

**Author(s)**

Beibei Jiang <beibei\_jiang@psych.mpg.de>

**Examples**

```
calc.gene.var(gene.coefficients, allele.freq)
```

---

calc.herit                      *Calculate heritability*

---

### Description

Calculate heritability

### Usage

```
calc.herit(gene.coef, freq, noise.var, Dskim = 0, ...)
```

### Arguments

gene.coef	a vector of 8 genetic parameters. Each element includes 8 genetic parameters (regression coefficient) $\beta_{Gw_t}$ , $t$ in $(1, 2, \dots, 8)$ .
freq	a dataframe with the allele frequencies.
noise.var	variance of noise to generate the random noise.
Dskim	the coefficient of linkage disequilibrium. Default is 0 (no LD).
...	not used.

### Value

heritability.

### Author(s)

Beibei Jiang <beibei\_jiang@psych.mpg.de>

---

check.snp.par                      *Check the number of the SNPs involved in epistasis and main effects*

---

### Description

Check the number of the SNPs set in the simulation parameters.

### Usage

```
check.snp.par(genetic.pars, nphe, ...)
```

### Arguments

genetic.pars	a data.frame or a matrix containing the parameter information for the main effect: additive and dominance.
nphe	number of phenotypes.
...	not used.

**Details**

The number of SNPs involved in main effects should be the same as the number of SNPs involved in epistasis.

**Value**

NULL — will stop if test fails.

**Author(s)**

Beibei Jiang <beibei\_jiang@psych.mpg.de>

**Examples**

```
check.snp.par(genepars, nphe = 2)
```

---

count.allele	<i>Count allele frequencies</i>
--------------	---------------------------------

---

**Description**

Count major and minor allele frequencies.

**Usage**

```
count.allele(x, ...)
```

**Arguments**

`x` a vector of single SNP information (minor allele count for genotype).  
`...` not used.

**Value**

a vector with major and minor allele frequency.

**Author(s)**

Beibei Jiang <beibei\_jiang@psych.mpg.de> and Benno Pütz <puetz@psych.mpg.de>

**Examples**

```
maf <- 0.1
x <- sample(0:2, 1000, replace = TRUE, prob = c((1-maf)^2, 2*(1-maf)*maf, maf^2))
table(x)
count.allele(x)
```

---

epistasis.pars      *Parameter settings of epistatic effects.*

---

**Description**

A dataset containing the parameter settings for epistatic effects. epistasis.pars.

**Usage**

```
data(epistasis.pars)
```

**Format**

A data frame with 6 rows and 3 columns(variables):

**SNPA** first SNP

**SNPB** second SNP

**additive\_additive** coefficient for additive-additive interaction

**additive\_dominance** coefficient for additive-dominance interaction

**dominance\_additive** coefficient for dominance-additive interaction

**dominance\_dominance** coefficient for dominance-dominance interaction

---

gene.coefficients      *Coefficients of genetic effects.*

---

**Description**

A dataset containing the regression coefficients of genetic effects. gene.coefficients.

**Usage**

```
data(gene.coefficients)
```

**Format**

A list with 3 elements:

**epi.par1** a data frame with 1 row and 10 variables:

**SNPA** first SNP

**SNPB** second SNP

**additiveA** coefficient for additive effect at locus A

**dominanceA** coefficient for dominance effect at locus A

**additiveB** coefficient for additive effect at locus B

**dominanceB** coefficient for dominance effect at locus B



**additive\_additive** coefficient for additive-additive interaction  
**additive\_dominance** coefficient for additive-dominance interaction  
**dominance\_additive** coefficient for dominance-additive interaction  
**dominance\_dominance** coefficient for dominance-dominance interaction

**epi.par2** a data frame with 1 row and 10 variables:

**SNPA** first SNP  
**SNPB** second SNP  
**additiveA** coefficient for additive effect at locus A  
**dominanceA** coefficient for dominance effect at locus A  
**additiveB** coefficient for additive effect at locus B  
**dominanceB** coefficient for dominance effect at locus B  
**additive\_additive** coefficient for additive-additive interaction  
**additive\_dominance** coefficient for additive-dominance interaction  
**dominance\_additive** coefficient for dominance-additive interaction  
**dominance\_dominance** coefficient for dominance-dominance interaction

**epi.par3** a data frame with 1 row and 10 variables:

**SNPA** first SNP  
**SNPB** second SNP  
**additiveA** coefficient for additive effect at locus A  
**dominanceA** coefficient for dominance effect at locus A  
**additiveB** coefficient for additive effect at locus B  
**dominanceB** coefficient for dominance effect at locus B  
**additive\_additive** coefficient for additive-additive interaction  
**additive\_dominance** coefficient for additive-dominance interaction  
**dominance\_additive** coefficient for dominance-additive interaction  
**dominance\_dominance** coefficient for dominance-dominance interaction

---

gene.effect

*Get genetic effect*

---

## Description

Get genetic effect for each individual based on the genotype.

## Usage

```
gene.effect(geno, gene.coef, model = c("epistasis"), ...)
```

## Arguments

geno	a data.frame or a matrix containing the genotype information.
gene.coef	a list with the coefficients of genetic effects.
model	a string specifying the genetic model to use for the simulation. Default is "epistasis".
...	not used.

**Value**

a data.frame including genetic effects.

**Author(s)**

Beibei Jiang <beibei\_jiang@psych.mpg.de> and Benno Pütz <puetz@psych.mpg.de>

**Examples**

```
# genotype file: rows are individuals and columns are SNPs
fgeno.path <- system.file("extdata", "10SNP.txt", package="SimPhe")

# get genotype
geno <- read.geno(fgeno.path, ftype = "snp.head")

# take a look at geno and gene.coef
geno
gene.coefficients

# get gene effects
gene.effect(geno, gene.coefficients)
```

---

genepars

*Parameter settings for simulation.*

---

**Description**

A dataset containing the parameters for a simulation. genepars.

**Usage**

```
data(genepars)
```

**Format**

A list with 7 elements:

**P1mean** A data frame with 1 row and 1 variable:

**mean**  $\beta_0$ : coefficient parameter of "basic" genetic effects in  $G_{ij} = \beta_0 + \sum_{t=1}^8 \beta_{G_{w_t}} w_{tij}$ .

**P1main** A data frame with 6 rows and 3 variables:

**SNP** SNP name

**additive** coefficient of additive effect

**dominance** coefficient of dominance effect

**P1epistasis** A data frame with 3 rows and 6 variables:

**SNPA** first SNP

**SNPB** second SNP

**additive\_additive** coefficient for additive-additive interaction  
**additive\_dominance** coefficient for additive-dominance interaction  
**dominance\_additive** coefficient for dominance-additive interaction  
**dominance\_dominance** coefficient for dominance-dominance interaction

**P1heritability** A data frame with 1 row and 1 variable:

**heritability** expected heritability

**P2mean** A data frame with 1 row and 1 variable:

**mean** mean of genetic effect

**P2main** A data frame with 6 rows and 3 variables:

**SNP** SNP name

**additive** coefficient of additive effect

**dominance** coefficient of dominance effect

**P2epistasis** A data frame with 3 rows and 6 variables:

**SNPA** first SNP

**SNPB** second SNP

**additive\_additive** coefficient for additive-additive interaction

**additive\_dominance** coefficient for additive-dominance interaction

**dominance\_additive** coefficient for dominance-additive interaction

**dominance\_dominance** coefficient for dominance-dominance interaction

---

genetic.scale

*Genetic scales of epistatic model*

---

## Description

Genetic scales of epistatic model (Cockerham model) based on F2 population.

## Usage

```
genetic.scale(SNPA = 0, SNPB = 0)
```

## Arguments

SNPA encoded alleles for first SNP.

SNPB encoded alleles for second SNP.

## Details

Calculate the genetic scale for a given allele combination of two SNPs. There are 9 genotypes in an F2 population, so we need 8 genetic parameters to give a complete description of the values for the 9 genotypes. Under the assumption of Hardy-Weinberg and linkage equilibrium, Cockerham (1954)'s orthogonal partition of genetic variance leads to the definition of the genotypic value  $G_{ij}$

$$G_{ij} = \beta_0 + \sum_{t=1}^8 \beta_{G_{w_t}} w_{tij}$$

by eight orthogonal scales or contrasts  $w_t$ 's,  $t$  in (1, 2, ..., 8). Four are marginal scales and four are interaction scales. Marginal scales (defined by Model I for an F2 population) are called linear and quadratic scales (additive and dominance scales in genetic terms). Correspondingly, the interaction scales are

**w1** additive for locus A;

**w2** dominance for locus A;

**w3** additive for locus B;

**w4** dominance for locus B;

**w5 (= w1 × w3)** linear × linear, additive × additive;

**w6 (= w1 × w4)** linear × quadratic, additive × dominance;

**w7 (= w2 × w3)** quadratic × linear, dominance × additive;

**w8 (= w2 × w4)** quadratic × quadratic, dominance × dominance.

SNPs are encoded by (0, 1, 2):

**0** means homozygous with major alleles;

**1** means heterozygote;

**2** means homozygous with minor alleles

E.g., the SNPs are encoded as 0: AA, 1: AG, and 2: GG, where 'A' represents the major allele and 'G' the minor allele.

## Value

a vector of genetic scales.

## Author(s)

Benno Pütz <puetz@mpipsyk1.mpg.de> and Beibei Jiang <beibei\_jiang@psych.mpg.de>

## Examples

```
genetic.scale(SNPA = 1, SNPB = 0)
```

---

get.freq	<i>Calculate the gene(allele) frequency</i>
----------	---

---

### Description

Calculate the gene(allele) frequency for each of the SNPs.

### Usage

```
get.freq(geno, epi.pars, ...)
```

### Arguments

geno	a dataframe of genotype data: columns are the SNPs; lines are individuals.
epi.pars	a data.frame or a matrix containing the parameter information for epistatic effect: additive $\times$ additive, additive $\times$ dominance, dominance $\times$ additive, and dominance $\times$ dominance.
...	not used

### Value

a dataframe with allele frequencies (major and minor).

### Author(s)

Beibei Jiang <beibei\_jiang@psych.mpg.de>

### Examples

```
# genotype file: rows are individuals and columns are SNPs
fgeno.path <- system.file("extdata", "10SNP.txt", package="SimPhe")

# get genotype
geno <- read.geno(fgeno.path, ftype = "snp.head")

get.freq(geno, epistasis.pars)
```

---

`get.gene.coef`*Get the coefficients of genetic effect*

---

**Description**

Get the coefficients of genetic effectsre.

**Usage**

```
get.gene.coef(main.pars, epi.pars, model = c("epistasis"), ...)
```

**Arguments**

<code>main.pars</code>	a data.frame or a matrix containing the parameters for the main effect: additive and dominance.
<code>epi.pars</code>	a data.frame or a matrix containing the parameters for the epistatic effect: additive $\times$ additive, additive $\times$ dominance, dominance $\times$ additive, dominance $\times$ dominance.
<code>model</code>	a string show the genetic model to use for simulation. Default is "epistasis"
<code>...</code>	not used.

**Value**

a list with the coefficients of genetic effects.

**Author(s)**

Beibei Jiang <beibei\_jiang@psych.mpg.de> and Benno Pütz <puetz@psych.mpg.de>

**Examples**

```
# take a look at the settings of coefficients for main effects
maineff.pars

# take a look at the settings of coefficients for interactive effects
epistasis.pars

# get a vector of gene coefficients
get.gene.coef(maineff.pars, epistasis.pars)
```

---

get.noise.var	<i>Suggestion noise</i>
---------------	-------------------------

---

**Description**

Give suggestion on the parameter setting of noise variance according to the expected heritability.

**Usage**

```
get.noise.var(gene.coef, freq, heritability, Dskim = 0, ...)
```

**Arguments**

gene.coef	a list including the coefficients of genetic effects. Each element includes 8 genetic parameters (regression coefficient) $\beta_{Gw_t}$ , $t$ in (1, 2, ..., 8)
freq	a dataframe with the allele frequencies.
heritability	expected heritability.
Dskim	the coefficient of linkage disequilibrium. Default is 0 (no LD).
...	not used.

**Value**

variance of noise to generate the random noise.

**Author(s)**

Beibei Jiang <beibei\_jiang@psych.mpg.de>

**Examples**

```
get.noise.var(gene.coefficients, allele.freq, 0.5)
```

---

list2frame	<i>Convert list to data.frame</i>
------------	-----------------------------------

---

**Description**

Convert list to data.frame.

**Usage**

```
list2frame(x, ...)
```

**Arguments**

`x` a list. In this package, it is used as a list includes simulated phenotypes. One element per block. Each element is a dataframe with SNP names and individuals.

`...` not used.

**Value**

a data.frame.

**Author(s)**

Beibei Jiang <beibei\_jiang@psych.mpg.de>

**Examples**

```
x <- list(test1=matrix(rnorm(1000), ncol=2), test2=matrix(rnorm(1000), ncol=2))
str(x)
x.new <- list2frame(x)
str(x)
```

---

maineff.pars

*Parameter settings of main effects (additive effect and dominance).*

---

**Description**

A dataset containing the parameter settings for main effects. maineff.pars.

**Usage**

```
data(maineff.pars)
```

**Format**

A data frame with 6 rows and 3 columns(variables):

**SNP** name of SNP

**additive** coefficient of additive effect

**dominance** coefficient of dominance effect



---

pars.writer                      *Write current parameters to file*

---

### Description

Write out the information about the parameters for simulation.

### Usage

```
pars.writer(genetic.pars, fname = "usedpars.txt", ...)
```

### Arguments

genetic.pars	list of simulation parameters. One element per block of file. Each element is a dataframe with SNP names and model parameters.
fname	filename of the setting for simulation (for recording).
...	not used.

### Author(s)

Beibei Jiang <beibei\_jiang@psych.mpg.de>

### Examples

```
pars.writer(genepars)
```

---

phe.writer                      *Write phenotypes*

---

### Description

Write out the simulated phenotypes.

### Usage

```
phe.writer(phe, onefile = TRUE, fname = "simu.pheno", ...)
```

### Arguments

phe	a data.frame or a matrix of simulated phenotypes. Each column is a phenotype.
onefile	whether to create just one file for all phenotypes (default) or one per phenotype
fname	filename of the phenotype(s).
...	not used.

**Author(s)**

Beibei Jiang <beibei\_jiang@psych.mpg.de> and Benno Pütz <puetz@psych.mpg.de>

**Examples**

```
phe <- matrix(rnorm(1000), ncol = 2)
colnames(phe) <- c("p1", "p2")
phe.writer(phe)
```

---

read.geno

*Read genotype data based on the file format*

---

**Description**

Read genotype data.

**Usage**

```
read.geno(fname = NULL, verbose = getOption("verbose"), run = TRUE,
  cleanup = TRUE, ftype = c("ind.head", "plink", "snp.head"),
  plink.path = NULL, ...)
```

**Arguments**

fname	a string specifying the file to read genotype information from
verbose	when set show the commands that are to be called through system.
run	when set (default) execute the system calls.
cleanup	when set (default) remove intermediate files before returning.
ftype	genotype file format, it accepts three options: <b>"plink"</b> : plink format (.bed, .bim, .fam or .map, .ped); <b>"ind.head"</b> : columns are the individuals and lines are SNPs; <b>"snp.head"</b> : columns are SNPs and lines are individuals.

For "plink", fgeno needs to be given without suffix and plink.path may need to be assigned by the user because plink will be run from within **SimPhe**. More detail see plink.path. For the other options, fgeno should be the full name (with suffix and path if necessary) of the genotype file. Of course, this does not apply if fgeno is provided as a data frame.

plink.path	path of plink executable. Only needed when the ftype is "plink". Default is NULL. The function will detect the plink path with system("where plink") for Windows users and system("which plink") for Linux and MacOS users. But there is no guarantee that the commands work on all devices. If the path cannot be determined or the executable cannot be called from read.geno, then users have to try other formats.
...	not used.

**Details**

If it is plink file format (.bed, .bim, .fam), make sure that plink has already been installed in the system

**Value**

a dataframe of genotype data: columns are the SNPs; rows are individuals.

**Author(s)**

Beibei Jiang <beibei\_jiang@psych.mpg.de> and Benno Pütz <puetz@psych.mpg.de>

**Examples**

```
## "snp.head" genotype file: rows are individuals and columns are SNPs
# get full path of example file
fgeno.path <- system.file("extdata", "10SNP.txt", package="SimPhe")

geno <- read.geno(fgeno.path, ftype = "snp.head")
head(geno)

## "plink" genotype file: 1).map and .ped; 2).bed, .fam and .bim
# get directory of plink example file
fpath <- strsplit(fgeno.path, "10SNP.txt")

#### Note: before run this example, specify your installation path of plink ####
# geno <- read.geno(paste0(fpath, "bdemo"), ftype = "plink", plink.path = "user's plink path")
```

---

read.simu.pars

*Read parameters*


---

**Description**

Read file specifying the simulation parameters.

**Usage**

```
read.simu.pars(file = NULL, ...)
```

**Arguments**

file	the file of parameters settings
...	not used

## Details

File format: please follow the example of the `simupars.txt` file found in the `inst/extdata/` directory of the package (run `system.file("extdata", "simupars.txt", package="SimPhe")` to see the full path), blank lines are ignored. The file consists of three or four blocks for each phenotype: mean, main, epistasis, and (optionally) heritability. Each block is started by a line of the form '[blockname]' followed by the parameters for the block, e.g., for the first phenotype,

### [P1mean ]

**mean**  $\beta_0$ : coefficient parameter of "basic" genetic effects in  $G_{ij} = \beta_0 + \sum_{t=1}^8 \beta_{G_{w_t}} w_{tij}$ .

### [P1main ]

**SNP** SNP name

**additive** coefficient of additive effect

**dominance** coefficient of dominance effect

### [P1epistasis ]

**SNPA** first SNP name

**SNPB** second SNP name

**additive\_additive** coefficient for additive-additive interaction

**additive\_dominance** coefficient for additive-dominance interaction

**dominance\_additive** coefficient for dominance-additive interaction

**dominance\_dominance** coefficient for dominance-dominance interaction

### [P1heritability ]

**heritability** expected heritability

For each block the expected columns and the respective meanings are given. Similar blocks need to be provided for the other phenotype(s): "[P2mean]", "[P2main]", "[P2epistasis]", and so on.

## Value

a list of simulation parameters. One element per block of file. Each element is a dataframe with SNP names and model parameters

## Author(s)

Beibei Jiang <beibei\_jiang@psych.mpg.de> and Benno Pütz <puetz@psych.mpg.de>

## Examples

```
# simulation parameters:
fpar.path <- system.file("extdata", "simupars.txt", package="SimPhe")

# pars <- read.simu.pars(fpar.path)
```

---

`regextract`*Extract (sub)strings matching regex pattern*

---

**Description**

Extract (sub)strings matching regex pattern.

**Usage**

```
regextract(x, pattern, ...)
```

**Arguments**

<code>x</code>	a character vector.
<code>pattern</code>	regular expression to be found in <code>x</code> .
<code>...</code>	not used.

**Details**

Extract the substrings of `x` that match the regex pattern `pattern`. The pattern may contain groups (enclosed in parentheses) which will result in further substrings extracted.

Derived from the help on [regmatches](#), take a look at the help there

**Value**

a character (string) matrix where the first column contains the global match for the pattern, each pair of `'()`' will result in another column with the respective match.

**Author(s)**

Benno Pütz <puetz@psych.mpg.de>

**Examples**

```
s <- "Test: A1 BC23 DEF456"
pattern = "[[:alpha:]]+([[:digit:]]+)"
regextract(s, pattern)

# equivalent to this example from the help page for grep()
lapply(regmatches(s, gregexpr(pattern, s)), function(e) regmatches(e, regexec(pattern, e)))
```

sim.phe

*Simulation for phenotypes (SimPhe main process)***Description**

Main process of simulation for phenotypes

**Usage**

```
sim.phe(sim.pars = NULL, fgeno = NULL, ftype = c("ind.head", "plink",
  "snp.head"), fwrite = TRUE, fphename = "simu.pheno",
  fusepar = "usedpars.txt", seed = NA, Dskim = 0, noise.var = 1,
  pattern = "[[:alpha:]]+", plink.path = system("which plink"),
  genetic.model = "epistasis", ...)
```

**Arguments**

**sim.pars** a prepared list containing the parameters settings for simulation or a file of parameters settings. Please set your own parameters following the same structure as the object `genepars` or as the file `simupars.txt` (you could find example file `system.file("extdata", "simupars.txt", package="SimPhe")`).

To specify heritability, there are two ways: one is to set heritability in parameter file or in the prepared list object which will further pass to `sim.pars`. Another way is to set `noise.var` by using function `get.noise.var` given specify heritability.

List format: please follow the example of the object `genepars`. The meaning of each element in the list is similar like the file format description below.

File format: please follow the example of the `simupars.txt` file found in the `inst/extdata/` directory of the package (run `system.file("extdata", "simupars.txt", package="SimPhe")` to get the path to the file), blank lines are ignored. The file consists of three or four blocks for each phenotype (the number of blocks depends on user): mean, main and epistasis, sometimes heritability. Each block is started by a line of the form '[blockname]' followed by the parameter setting for the block, e.g. for first phenotype,

**[P1mean ]**

**mean**  $\beta_0$ : coefficient parameter of "basic" genetic effects in  $G_{ij} = \beta_0 + \sum_{t=1}^8 \beta_{G_{w_t}} w_{tij}$ .

**[P1main ]**

**SNP** SNP name

**additive** coefficient of additive effect

	<b>dominance</b> coefficient of dominance effect
	<b>[P1epistasis ]</b>
	<b>SNPA</b> first SNP
	<b>SNPB</b> second SNP
	<b>additive_additive</b> coefficient for additive-additive interaction
	<b>additive_dominance</b> coefficient for additive-dominance interaction
	<b>dominance_additive</b> coefficient for dominance-additive interaction
	<b>dominance_dominance</b> coefficient for dominance-dominance interaction
	<b>[P1heritability ]</b>
	<b>heritability</b> expected heritability
	Similar meanings for "[P2mean]", "[P2main]", "[P2epistasis]", and so on.
fgeno	file to read genotype information from or pre-read data.frame with that information (matching the output format of <a href="#">read.geno</a> ).
ftype	genotype file format, it accepts three options: <b>"plink"</b> : plink format (.bed, .bim, .fam or .map, .ped); <b>"ind.head"</b> : columns are the individuals and lines are SNPs; <b>"snp.head"</b> : columns are SNPs and lines are individuals. For "plink", fgeno needs to be given without suffix and plink.path may need to be assigned by the user because plink will be run from within <b>SimPhe</b> . More detail see plink.path. For the other options, fgeno should be the full name (with suffix and path if necessary) of the genotype file. Of course, this does not apply if fgeno is provided as a data frame.
fwrite	logical. Write out file (simulated data) or not. If TRUE (default), simulated phenotypes will be written, respectively.
fphename	filename of the phenotype(s). Default is "simu.pheno".
fusepar	filename of the setting for simulation (for recording). Default is "usedpars.txt".
seed	an integer used for set.seed(). Default is NA.
Dskim	the coefficient of linkage disequilibrium. Default is 0 (no LD).
noise.var	variance for random noise. Default is 1. Note that this is overridden by the heritability setting in the simulation parameter file. If heritability is given in parameter file then noise.var will not work.
pattern	ignore pattern for detecting the phenotype index from the parameter names. Default is "[[:alpha:]]+" which means letters.
plink.path	path of plink executable. Only needed when the ftype is "plink". Default is NULL. The function will detect the plink path with system("where plink") for Windows users and system("which plink") for Linux and MacOS users. But there is no guarantee that the commands work on all devices. If the path cannot be determined or the executable cannot be called from read.geno, then users have to try other formats.
genetic.model	a string show the genetic model to use for simulation. Default is "epistasis".
...	not used.

**Details**

further discussion on pattern

**Value**

a data.frame with the simulated phenotype(s) where the column(s) refer to different phenotype(s) and rows to individuals.

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**Examples**

```
#### file path of example
# simulation parameters:
fpar.path <- system.file("extdata", "simupars.txt", package="SimPhe")

# genotype file: rows are individuals and columns are SNPs
fgeno.path <- system.file("extdata", "10SNP.txt", package="SimPhe")

#### instead of a parameter file, prepared list like genepars also works
genepars

#### simulate phenotype(s)
phe <- sim.phe(sim.pars = fpar.path, fgeno = fgeno.path, ftype = "snp.head", fwrite = FALSE)
# or
phe <- sim.phe(sim.pars = genepars, fgeno = fgeno.path, ftype = "snp.head", fwrite = FALSE)

# the simulated phenotype(s)
str(phe)
head(phe)
```

---

specify.pars

*Get the parameters of main/epistatic effects per phenotype*

---

**Description**

Get the parameters of main/epistatic effects per phenotype.

**Usage**

```
specify.pars(genetic.pars, effect.type = c("main", "epistasis"),
  phe.index = 1, ...)
```



**Arguments**

<code>genetic.pars</code>	a data.frame or a matrix containing the parameters information for main effect: additive and dominance.
<code>effect.type</code>	a string naming the type of the genetic effects (accepts either "main" or "epistasis").
<code>phe.index</code>	a integer indicating the phenotype. Default is 1.
<code>...</code>	not used.

**Value**

a data.frame or a matrix containing the parameters information for epistatic effect: additive  $\times$  additive, additive  $\times$  dominance, dominance  $\times$  additive, dominance  $\times$  dominance.

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**Examples**

```
# get parameters of coefficients for main effects
specify.pars(genepars, effect.type = "main")

# get parameters of coefficients for interactive effects
specify.pars(genepars, effect.type = "epistasis")
```

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